



Technology Transition Workshop | *Kristin S. Lowery, Ph.D.*

Ibis™ Assay Workflow Overview

Ibis™ Assay Workflow

- 1. Receive and store assay kit components.**
- 2. Perform up-front sample processing.**
- 3. Register the experiment.**
- 4. Set up PCR plate(s).**
- 5. Seal the PCR plates.**
- 6. Thermocycle PCR plates.**
- 7. Prepare reagents for PLEX-ID™.**
- 8. Fill reagents on PLEX-ID™ and empty waste.**
- 9. Analyze the PCR plates on the PLEX-ID™.**
- 10. Review data.**

Ibis™ Assay Workflow – Step 1

- **Upon receipt, check for assay components and store as indicated**
 - **10 barcoded assay plates**
 - **Store @ -20° C – manual defrost freezer**
 - **1 bottle of magnetic beads**
 - **Store @ 4° C and in upright position**
 - **1 reservoir for magnetic beads**
 - **Store @ room temperature**
 - **3 cleanup reagents (CR1, CR2 and CR3)**
 - **Store @ 4° C**
 - **Instructions sheet**

Ibis™ Assay Workflow – Step 2

- **Up-front sample processing**
- **Extract DNA from samples (your choice of method)**
 - **Qiagen® columns**
 - **KingFisher® magnetic bead systems**
 - **Phenol / chloroform**
 - **Others**

Ibis™ Assay Workflow – Step 3


- **Register experiment**
 - For data analysis to work correctly, the plate must contain a positive control and a negative control
 - 10 samples can be run on the plate
 - Use different wizards depending on number of samples
 - Requires sample list – Microsoft® Excel spreadsheet
 - Can use tubes or plates for setup
 - Use control layouts
 - Define positions for negative and positive PCR controls
 - Do not need to put controls in sample list

Ibis™ Assay Workflow – Step 4

- Set up PCR plates either by hand or on reformatting robot
- 5 μL of sample is added per well
 - A total of 50 μL is needed for each sample



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Ibis™ Assay Workflow – Step 5



Image courtesy of <http://www.thermo.com/com/cda/product/detail/1,,10142764,00.html>

- Heat seal plate
- Centrifuge

Ibis™ Assay Workflow – Step 6

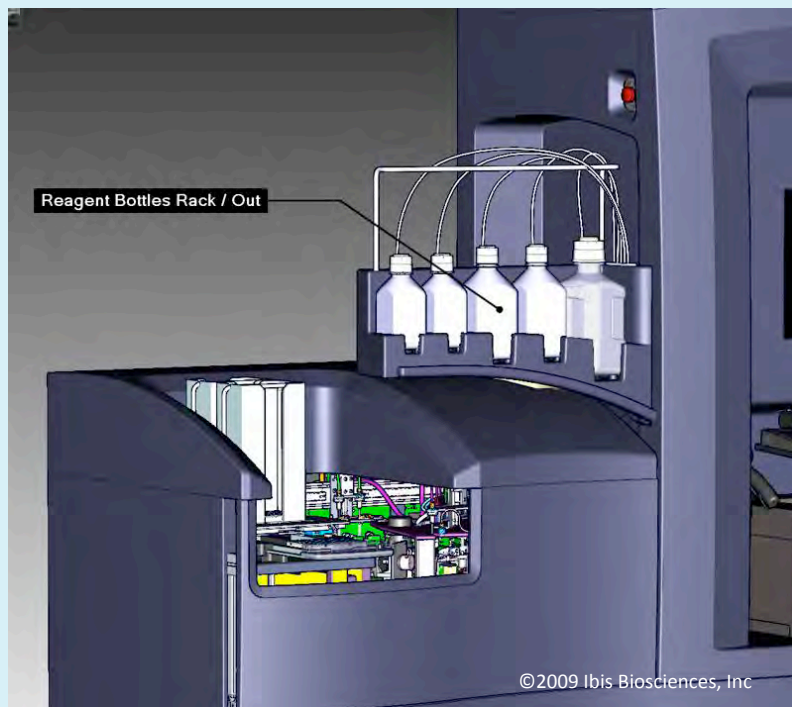


- Thermocycle the PCR plates
- Programs contain 10 minute step at 99° C to minimize enzyme activity
- After thermocycling, centrifuge the plate for ~15 seconds at ~800 rpm
- Plate may be frozen until put on PLEX-ID™

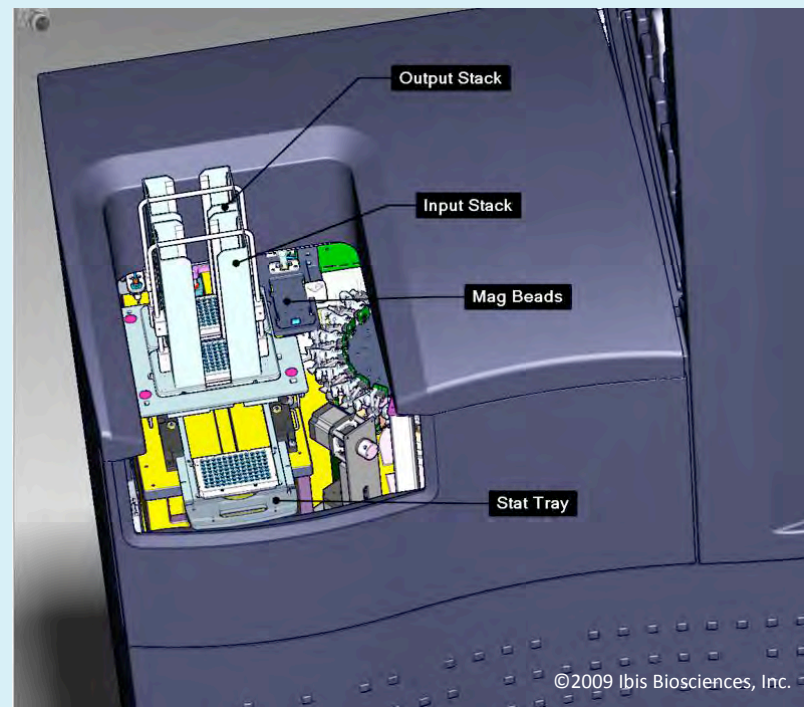
Ibis™ Assay Workflow – Step 7

- **Prepare magnetic bead reservoir**
 - Dispense beads into reservoir
 - Seal with tape provided
- **Prepare clean-up reagents**
 - Add volume of methanol indicated on bottle
 - Use Burdick and Jackson™ HPLC grade
- **Recommend tracking lot numbers of reagents**

Ibis™ Assay Workflow – Step 8



Fill reagents and empty waste



Insert magnetic bead reservoir and plates

Ibis™ Assay Workflow – Step 9



Run PCR plates on the PLEX-ID™

Ibis™ Assay Workflow – Step 10

- View results in IbisTrack

The screenshot shows the IbisTrack software interface. The main window displays the 'View Results' step for a specific assay. The interface includes a menu on the left with options like 'Register Assays', 'Manage Inventory', and 'STR Analysis'. The main window displays a plate layout for 'Plate P05010295' with sample 12. It shows a 'Register' button and a 'Generate report for P05010295' button. The 'Analysis' tab is active, showing a 'P05010295-SC35495-10-POS composite' spectrum and two well-specific spectra: 'Well 12 (A12)' and 'Well 24 (B12)'. The spectrum displays peaks with their corresponding mass values (e.g., 2906, 2901, 2892 for Well 12). A table on the right lists peak data with columns for 'num', 'error', 'exp. mass', and 'obs. I'.

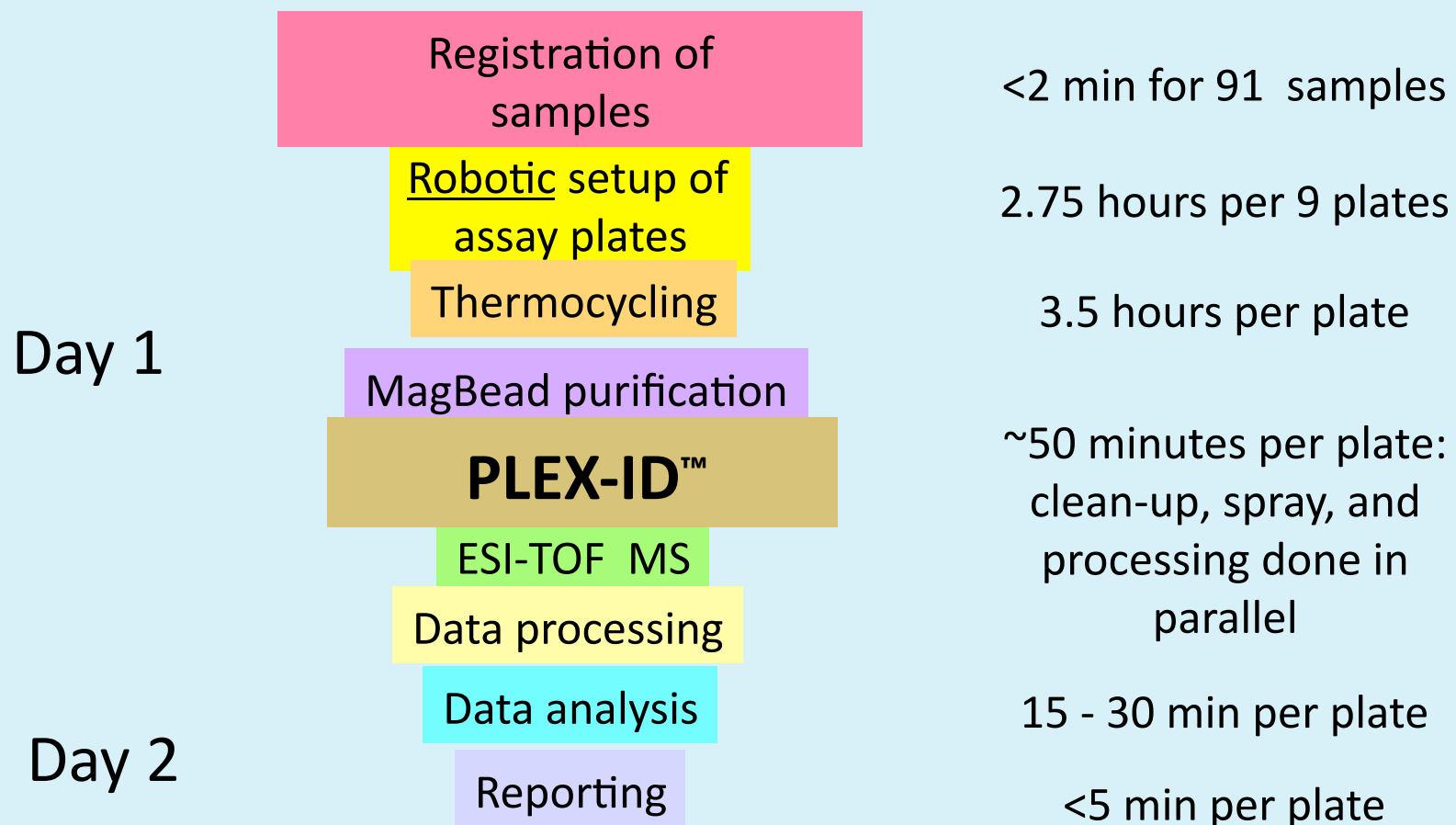
Workflow Timeline

- **Sample Prep**
 - Depending on method 30 to 60 minutes
- **Plate Setup**
 - Manual: 10 to 20 minutes per plate
 - Reproducibility is an issue
 - Robotic: 15 minutes per plate
- **PCR**
 - PLEX-ID™ STR Assay – ~3 hours
 - PLEX-ID™ mtDNA Assay – 3.5 hours
 - PLEX-ID™ SNP Assay – 2.5 hours

Workflow Timeline

- **PLEX-ID™**
 - **Initial flushing and system startup: 20 minutes**
 - **Clean-up: 10 minutes for first well, then one well cleaned every 30 seconds**
 - **Spray on TOF: ~50 minutes per plate**
 - **30 seconds per well – 4 minutes per 8 well sample**
 - **Data processing: 15 to 20 minutes per plate**

Workflow Timeline



Workflow Throughput

- **Assuming manual PCR setup and 2 cyclers, 4 plates a day**
 - 40 samples per day
 - 200 samples per week
 - 10,400 samples per year
- **Assuming robotic PCR setup and 5 cyclers, 10 plates a day**
 - 100 samples per day
 - 500 samples per week
 - 26,000 samples per year
- **Limiting factor – number of thermocyclers**

Questions?

Contact Information

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Note: All images are courtesy of Dr. Kristin S. Lowery unless otherwise noted.